



The 65th ASH Annual Meeting Abstracts

POSTER ABSTRACTS

617.ACUTE MYELOID LEUKEMIAS: BIOMARKERS, MOLECULAR MARKERS AND MINIMAL RESIDUAL DISEASE IN DIAGNOSIS AND PROGNOSIS**PET Tracer-Based Glucose Uptake Predicts Clinical Response to Induction Therapy in Acute Myeloid Leukaemia**Hui Zeng, MDPhD¹, Suqi Deng², Juan Du², Huien Zhan², Kexiu Huang¹, Xue Zheng², Ying Xu¹, Lu Wang³¹The First Affiliated Hospital of Jinan University, Guangzhou, China²The First Affiliated Hospital of Jinan University, Guangzhou, CHN³Department of Nuclear Medicine and PET/CT-MRI Center, The First Affiliated Hospital of Jinan University, Guangzhou, China

Metabolic re-programming enables cancer cells to harness various carbon substrates for energy production and biomass synthesis. Cell-intrinsic mechanisms of preferential glucose uptake is reported in acute myeloid leukaemia (AML). We previously identified leukaemia cells uptake more glucose compared with bone marrow micro-environment (BMME) cells in a MLL-AF9-driven AML mouse model (Leukemia 2023;37:1407). It is unknown how glucose is partitioned in the bone marrow, and whether glucose uptake variation correlates with response to induction chemotherapy in persons with newly-diagnosed AML. The radionuclide-labeled glucose analog ¹⁸F-2'-deoxy-2'-fluorodeoxyglucose (¹⁸F-FDG), a positron emission tomography (PET) probe, can be used to quantify in vivo glucose uptake in this setting. We studied this via a prospective pilot study (NCT05919199) in subjects with newly-diagnosed AML before receiving induction chemotherapy.

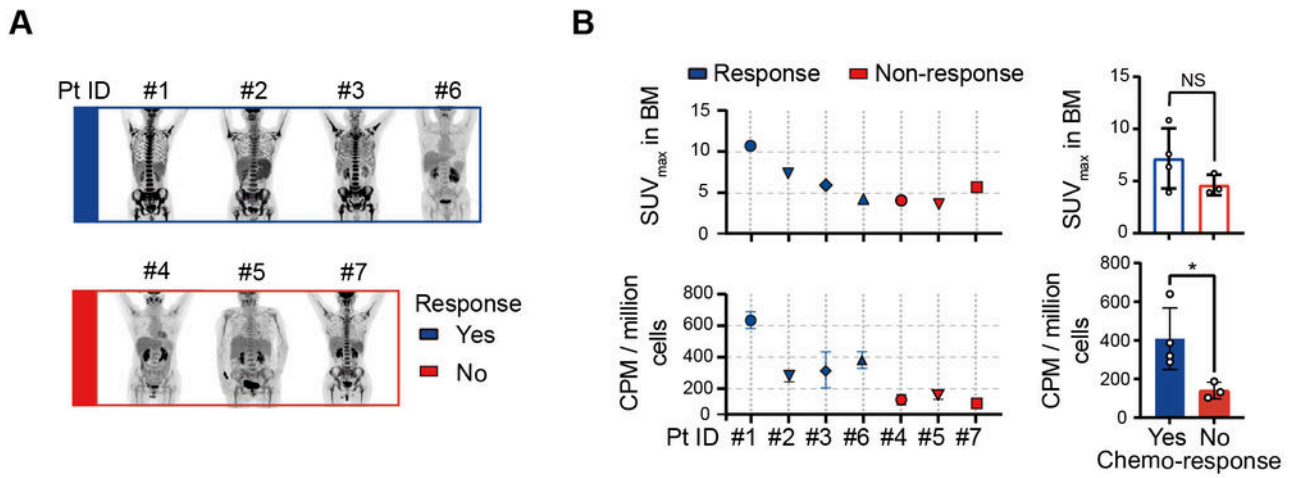
Of 7 patients who have been enrolled in the study to date, all subjects were presented with various intensities of diffuse bone marrow ¹⁸F-FDG uptake in PET/CT images (maximum standardized uptake value (SUV_{max}) mean, 6.0; range, 3.8-10.6). Subjects with higher SUV_{max} in bone marrow had a higher likelihood of achieving a remission after induction therapy (Figure A).

We use a ¹⁸F-FDG probe combined with magnetic-activated cell sorting (MACS) to quantify glucose uptake (¹⁸F radioactivity) in bulk and/or fractionated cell populations from subjects with AML at diagnosis. The associations between ¹⁸F-FDG uptake and response was better defined by per-cell glucose uptake quantification (¹⁸F counts per minute (CPM) per million cells) of bulk bone marrow cells. Higher ¹⁸F-FDG uptake correlated with a higher remission rate (Figure B). Notably, leukaemia cells from responders metabolized more glucose compared with micro-environment cells both of which had higher glucose uptake compared with non-responders. In contrast, leukaemia cells from non-responders had markedly decreased glucose uptake compared with micro-environment cells even they were sharing the same metabolic pool. These data indicate subject-specific glucose uptake is not directly affected by glucose availability but by cell-intrinsic glucose uptake programs.

In light of data implicating mitochondrial metabolism in preserving leukaemia stem cells we speculated other energy sources such as amino acids and, fatty acids might compensate for decreased glucose uptake in non-responders. We utilized a computationally metabolic fluxome model, scFEA (Genome Res 2021;31:1867), which characterizes stoichiometric relationships of metabolic networks and principal component analyses based on metabolism-related transcripts clustered samples by response. scFEA analysis of RNA-seq indicated responders had higher flux in the glucose import module, while flux of glutamine uptake were enriched in non-responders. These results suggest a switch from glucose ("Hot" AML) to glutamine metabolism ("Cold" AML) may protect leukaemia cells from chemotherapy stress.

Taken together, we provide a rationale for in vivo evaluation of glucose uptake in AML patients, which is of potential use in discriminating patient response or resistance to induction therapies.

Disclosures No relevant conflicts of interest to declare.



(A) Maximum intensity projection (MIP) from AML patients underwent ¹⁸F-FDG PET /CT-scanned at diagnosis. (B) Maximum SUV detected in bone marrow by PET/CT (upper panel); CPM per million cells in bulk BMNCs (lower panel).

Figure 1

<https://doi.org/10.1182/blood-2023-189148>